

We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

4,800

Open access books available

122,000

International authors and editors

135M

Downloads

Our authors are among the

154

Countries delivered to

TOP 1%

most cited scientists

12.2%

Contributors from top 500 universities



WEB OF SCIENCE™

Selection of our books indexed in the Book Citation Index
in Web of Science™ Core Collection (BKCI)

Interested in publishing with us?
Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected.
For more information visit www.intechopen.com



Recent Advances in Stem Cell and Tissue Engineering

Farideh Mohammadian

Additional information is available at the end of the chapter

<http://dx.doi.org/10.5772/intechopen.75967>

Abstract

The clinical application of stem cells in tissue engineering and regeneration is becoming more significant. However, its application has been limited by issues like reproducibility of the stem cells, ethical concerns of harvesting some of these stem cells, and controlling the fate of stem cells in vitro and in vivo. The advent of tissue engineering and regeneration has led to the fabrication of advanced biomaterials and scaffolds with enhanced ability to mimic and control the cellular microenvironment similar to that of innate stem cell niche. Combining the use of stem cells with biomaterials and scaffolds especially synthetic hydrogels that have exhibited physicochemical abilities and properties similar to native niche can be the future of tissue engineering in terms of formation of new tissues like bones. Recently, there has been an increase in the use of either endothelial progenitor cells (EPCs), induced pluripotent stem cells (iPSCs), or adult mesenchymal stem cells in preclinical studies; however this is yet to be transferred to clinical setups as there are limitations in terms of regulations and ethical considerations. The purpose of this review is to give comprehensive details about the application of stem cells in tissue engineering.

Keywords: endothelial progenitor cells, induced pluripotent stem cells, adult mesenchymal stem, tissue engineering, scaffolds

1. Introduction

Tissue engineering is a multidisciplinary science that applies the principles of bioengineering for the fabrication of new and improved biomaterials capable of maintaining and restoring the functionality of organs and tissues impaired by disease and trauma. This translational approach has been applied to develop and design patient-specific tissue grafts that mimic the functional properties of native tissues. Three important factors have been accredited to the success of tissue engineering: cocultured stem cells, signaling factor, and the bio-fabricated scaffold.

The stem cells are capable of differentiating into several types of tissues and organs, while the bio-fabricated scaffold provides structural support to the seeded stem cells. Signaling factors are responsible for influencing cell phenotype, metabolism, migration, and organization.

Stem cells are undifferentiated cells of embryonic, fetal origin, and they possess the ability to give rise to differentiated cells and then finally develop into organs. Stem cell characteristics include the ability to self-replicate and renew, clonage forming, and high potency ability [1]. In terms of the potency ability of stem cells, stem cells can be totipotent, could differentiate into any cell types (pluripotent) [2], and could differentiate into cells that arise from the three germ layers—ectoderm, endoderm, and mesoderm—from which organs develop [3].

Stem cells can be categorized broadly into embryonic and adult stem cells and are efficient cell sources for tissue regenerative applications. They have also been reported to have the abilities to promote tissue homeostasis, growth, and repair, thereby contributing importantly to tissue and organ regeneration [4]. Bio-fabricated scaffolds consist of decellularized biomaterials to provide structural and anatomical functions to the seeded stem cells, thereby resulting into successful formation of specific tissue. In support of the above report, Kang and colleagues demonstrated that decellularized scaffolds loaded with autologous adipose-derived stem cells (ADSCs) were efficient to repair cartilage defect in an animal model [5]. They concluded that decellularized scaffolds loaded with ADSC induced significant and improved cartilage tissue repair compared to native cartilage.

2. Mesenchymal stem cells seeded for bone tissue engineering

MSCs are stromal stem cells that are heterogeneous and are derived from several tissue sources that include adipose tissue [6], periodontal ligaments [7], bone marrow (**Figure 1**) [8], umbilical cord (UC) [9], placenta [10], and lungs [11]. MSCs express surface markers like CD73, CD44, CD90, and CD105. The most widely known and used MSCs are bone marrow MSCs and adipose tissue-derived MSCs isolated and purified from the bone marrow and adipose tissue, respectively. Briefly, the anatomy of the bone marrow is made up of the parenchyma and the stroma part. The parenchyma houses the hematopoietic stem cells, and the stroma part consists of the bone marrow stromal cells (MSCs) that have the capability to differentiate into several cell lines like osteoblasts, chondrocytes, adipocytes, etc. The clinical use of both bone-derived mesenchymal stem cells and adipose stem cells in bone tissue engineering has been reported using various models of bone regeneration such as osteogenesis [12, 13], long bone defects [14, 15], and calvarial defects [16, 17]. Furthermore, co-administration of stem cells with cytokines has been reported to be efficient in bone repair as cytokines and growth factors like stromal derived factor-1 (SDF-1) can lead to increased migration and homing of stem cells to the defected site [18]. In a similar report by Ho et al., they demonstrated that co-administration of stromal-derived factor-1 with BM-MSCs would indirectly enhance bone repair by improving migration of innate cells to the site of bone fracture. They concluded that BM-MSCs overexpressing SDF-1 were efficient in improving the new bone formation during the early stage of fracture healing compared to BM-MSC treatment alone [19]. Genes implicated in fracture healing such as osterix [20], hypoxia-inducible factor-1 [21], and BMP-7 [22] have all been reported to be efficient in bone formation when transfected with MSCs.

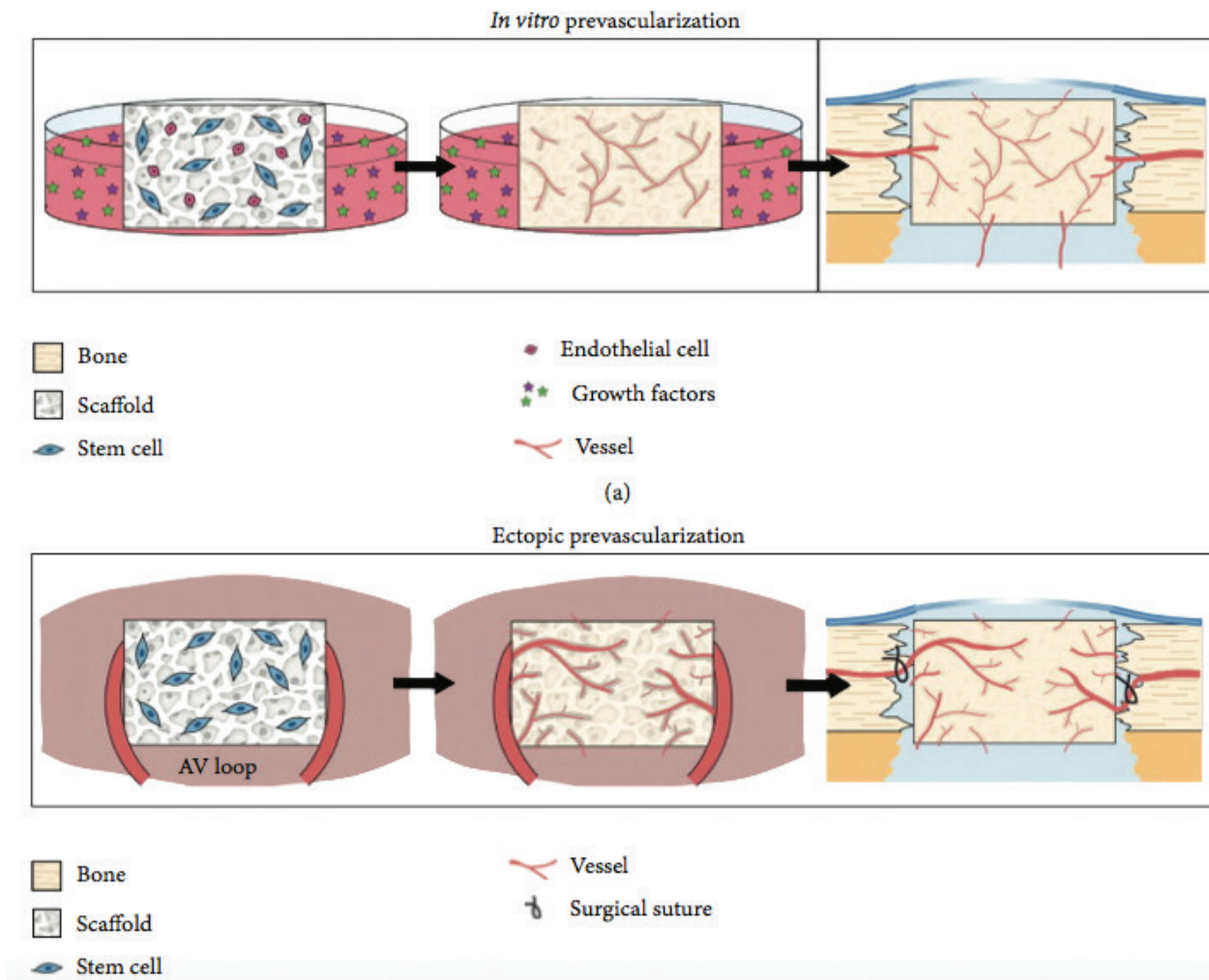


Figure 1. Showing in vivo and in vitro stem cell application in engineered tissue (a) *In vitro* prevascularization methods induce cell-seeded scaffolds to form vasculature (b) *In vivo* ectopic prevascularization involves implantation of a cell-seeded scaffold into a highly vascularized bed. Adapted from [26] with copyright permission.

3. Advances in MSCs and tissue engineering technology

Recently, bone tissue engineering in combination with novel stem cell-based technologies is yielding promising results as reported by Syed-Picard and colleagues in their experimental study that BM-MSC-derived cell sheets could be used to fabricate functional periosteal tissue [23]. Briefly, culturing BM-MSCs to hyperconfluence to produce abundant extracellular matrix to form robust cell sheets generated the BM-MSC-derived cell sheets. The authors reported that the generated cell sheets supported with calcium phosphate pellets were transplanted subcutaneously into mice for 8 weeks. They concluded that there was significant bone-like tissue formation by the BM-MSC-calcium phosphate pellet structure compared to the non-seeded calcium phosphate scaffold.

In another similar study by [24], BM-MSC cell sheet technology was compared to control cell complex. The authors reported that BM-MSC cell sheet resulted into significant expressed levels of growth factors crucial to bone development like vascular endothelial growth factor and PDGF. In another innovative study of stem cell application in tissue engineering, Ren et al. fabricated

- [29] Kaigler D et al. Stem cell therapy for craniofacial bone regeneration: A randomized, controlled feasibility trial. *Cell Transplantation*. 2013;**22**(5):767-777
- [30] Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell*. 2006;**126**(4):663-676
- [31] Ardeshtyrlajimi A, Dinarvand P, Seyedjafari E, Langroudi L, Jamshidi Adegani F, Soleimani M. Enhanced reconstruction of rat calvarial defects achieved by plasma-treated electrospun scaffolds and induced pluripotent stem cells. *Cell and Tissue Research*. 2013;**354**(3):849-860
- [32] Liu J, Chen W, Zhao Z, Xu HHK. Effect of NELL1 gene overexpression in iPSC-MSCs seeded on calcium phosphate cement. *Acta Biomaterialia*. 2014;**10**(12):5128-5138
- [33] Kang H et al. Mineralized gelatin methacrylate-based matrices induce osteogenic differentiation of human induced pluripotent stem cells. *Acta Biomaterialia*. 2014;**10**(12):4961-4970
- [34] Lian Q et al. Functional mesenchymal stem cells derived from human induced pluripotent stem cells attenuate limb ischemia in mice. *Circulation*. 2010;**121**(9):1113-1123
- [35] Atesok K et al. An emerging cell-based strategy in orthopaedics: Endothelial progenitor cells. *Knee Surgery, Sports Traumatology, Arthroscopy*. 2012;**20**(7):1366-1377
- [36] Asahara T. Isolation of putative progenitor endothelial cells for angiogenesis. *Science*. 1997;**275**(5302):964-966
- [37] Zigdon-Giladi H, Michaeli-Geller G, Bick T, Lewinson D, Machtei EE. Human blood-derived endothelial progenitor cells augment vasculogenesis and osteogenesis. *Journal of Clinical Periodontology*. 2015;**42**(1):89-95
- [38] Li R et al. Endothelial progenitor cells for fracture healing: A microcomputed tomography and biomechanical analysis. *Journal of Orthopaedic Trauma*. 2011;**25**(8):467-471
- [39] Kuroda R et al. Local transplantation of G-CSF-mobilized CD34 + cells in a patient with tibial nonunion: A case report. *Cell Transplantation*. 2011;**20**(9):1491-1496
- [40] Moore MA et al. Decellularization of human dermis using non-denaturing anionic detergent and endonuclease: A review. *Cell and Tissue Banking*. 2015;**16**(2):249-259
- [41] Hung SH, Su CH, Lee FP, Tseng H. Larynx decellularization: Combining freeze-drying and sonication as an effective method. *Journal of Voice*. 2013;**27**(3):289-294
- [42] de Peppo GM et al. Engineering bone tissue substitutes from human induced pluripotent stem cells. *Proceedings of the National Academy of Sciences of the United States of America*. 2013;**110**(21):8680-8685
- [43] Fröhlich M, Grayson WL, Marolt D, Gimble JM, Kregar-Velikonja N, Vunjak-Novakovic G. Bone grafts engineered from human adipose-derived stem cells in perfusion bioreactor culture. *Tissue Engineering. Part A*. 2010;**16**(1):179-189

- [44] Vincentelli A et al. In vivo autologous recellularization of a tissue-engineered heart valve: Are bone marrow mesenchymal stem cells the best candidates? *The Journal of Thoracic and Cardiovascular Surgery*. 2007;**134**(2):424-432
- [45] Nichols JE et al. Production and assessment of Decellularized pig and human lung scaffolds. *Tissue Engineering. Part A*. 2013;**19**(17-18):2045-2062
- [46] Ross EA et al. Embryonic stem cells proliferate and differentiate when seeded into kidney scaffolds. *Journal of the American Society of Nephrology*. 2009;**20**(11):2338-2347
- [47] Baiguera S, Del Gaudio C, Kuevda E, Gonfiotti A, Bianco A, Macchiarini P. Dynamic decellularization and cross-linking of rat tracheal matrix. *Biomaterials*. 2014;**35**(24):6344-6350
- [48] Baiguera S et al. Electrospun gelatin scaffolds incorporating rat decellularized brain extracellular matrix for neural tissue engineering. *Biomaterials*. 2014;**35**(4):1205-1214
- [49] Gray FL, Turner CG, Ahmed A, Calvert CE, Zurakowski D, Fauza DO. Prenatal tracheal reconstruction with a hybrid amniotic mesenchymal stem cells-engineered construct derived from decellularized airway. *Journal of Pediatric Surgery*. 2012;**47**(6):1072-1078
- [50] Nagaoka Y, Yamada H, Kimura T, Kishida A, Fujisato T, Takakuda K. Reconstruction of small diameter arteries using decellularized vascular scaffolds. *Journal of Medical and Dental Sciences*. 2014;**61**(1):33-40
- [51] Bertanha M et al. Morphofunctional characterization of decellularized vena cava as tissue engineering scaffolds. *Experimental Cell Research*. 2014;**326**(1):103-111
- [52] Baiguera S et al. Tissue engineered human tracheas for in vivo implantation. *Biomaterials*. 2010;**31**(34):8931-8938
- [53] Pei M, Zhang Y, Li J, Chen D. Antioxidation of decellularized stem cell matrix promotes human synovium-derived stem cell-based chondrogenesis. *Stem Cells and Development*. 2013;**22**(6):889-900
- [54] Jiang WC, Cheng YH, Yen MH, Chang Y, Yang VW, Lee OK. Cryo-chemical decellularization of the whole liver for mesenchymal stem cells-based functional hepatic tissue engineering. *Biomaterials*. 2014;**35**(11):3607-3617
- [55] Rana D, Zreiqat H, Benkirane-Jessel N, Ramakrishna S, Ramalingam M. Development of decellularized scaffolds for stem cell-driven tissue engineering. *Journal of Tissue Engineering and Regenerative Medicine*. 2017;**11**(4):942-965
- [56] Oberwallner B et al. Preparation of cardiac extracellular matrix scaffolds by decellularization of human myocardium. *Journal of Biomedical Materials Research Part A*. 2014;**102**(9):3263-3272
- [57] Dohmen PM et al. Mid-term clinical results using a tissue-engineered pulmonary valve to reconstruct the right ventricular outflow tract during the Ross procedure. *The Annals of Thoracic Surgery*. 2007;**84**(3):729-736

- [58] Macchiarini P et al. Clinical transplantation of a tissue-engineered airway. *Lancet*. 2008; **372**(9655):2023-2030
- [59] Gonfiotti A et al. The first tissue-engineered airway transplantation: 5-year follow-up results. *Lancet*. 2014;**383**(9913):238-244
- [60] Elliott MJ et al. Stem-cell-based, tissue engineered tracheal replacement in a child: A 2-year follow-up study. *Lancet*. 2012;**380**(9846):994-1000
- [61] Berg M et al. Replacement of a tracheal stenosis with a tissue-engineered human trachea using autologous stem cells: A case report. *Tissue Engineering, Part A*. 2014;**20**(1-2): 389-397
- [62] Sterodimas A, de Faria J, Nicaretta B, and Pitanguy I. Tissue engineering with adipose-derived stem cells (ADSCs): Current and future applications. *Journal of Plastic, Reconstructive & Aesthetic Surgery*. 2010;**63**(11):1886-1892
- [63] Masuda T, Furue M, Matsuda T. Novel strategy for soft tissue augmentation based on transplantation of fragmented omentum and preadipocytes. *Tissue Engineering*. 2004; **10**(11-12):1672-1683
- [64] Matsumoto D et al. Cell-assisted lipotransfer: Supportive use of human adipose-derived cells for soft tissue augmentation with lipoinjection. *Tissue Engineering*. 2006;**12**(12): 3375-3382
- [65] a Moseley T, Zhu M, Hedrick MH. Adipose-derived stem and progenitor cells as fillers in plastic and reconstructive surgery. *Plastic and Reconstructive Surgery*. 2006;**118**: 121S-128S
- [66] Zhu M et al. Supplementation of fat grafts with adipose-derived regenerative cells improves long-term graft retention. *Annals of Plastic Surgery*. 2010;**64**(2):222-228
- [67] Rasmussen JG et al. Prolonged hypoxic culture and trypsinization increase the pro-angiogenic potential of human adipose tissue-derived stem cells. *Cytotherapy*. 2011; **13**(3):318-328
- [68] Rubina K et al. Adipose stromal cells stimulate angiogenesis via promoting progenitor cell differentiation, secretion of Angiogenic factors, and enhancing vessel maturation. *Tissue Engineering, Part A*. 2009;**15**(8):2039-2050
- [69] Coleman SR, Saboeiro AP. Fat grafting to the breast revisited: Safety and efficacy. *Plastic and Reconstructive Surgery*. 2007;**119**(3):775-785-787
- [70] Yoshimura K, Sato K, Aoi N, Kurita M, Hirohi T, Harii K. Cell-assisted lipotransfer for cosmetic breast augmentation: Supportive use of adipose-derived stem/stromal cells. *Aesthetic Plastic Surgery*. 2008;**32**(1):48-55
- [71] Yoshimura K et al. Progenitor-enriched adipose tissue transplantation as rescue for breast implant complications. *The Breast Journal*. 2010;**16**(2):169-175

- [72] Kitamura SKK, Kajitani K, Hedrick M. Stem cell augmented reconstruction: A new hope for reconstruction after breast conservation therapy. *Breast Cancer Research and Treatment*. 2011;**106**
- [73] Tissiani LAL, Alonso N. A prospective and controlled clinical trial on stromal vascular fraction enriched fat grafts in secondary breast reconstruction. *Stem Cells International*. 2016;**2016**
- [74] Claro F, Figueiredo JCA, Zampar AG, Pinto-Neto AM. Applicability and safety of autologous fat for reconstruction of the breast. *British Journal of Surgery*. 2012;**99**(6):768-780
- [75] Böttcher-Haberzeth S et al. Characterization of pigmented dermo-epidermal skin substitutes in a long-term in vivo assay. *Experimental Dermatology*. 2015;**24**(1):16-21
- [76] Trottier V, Marceau-Fortier G, Germain L, Vincent C, Fradette J. IFATS collection: Using human adipose-derived stem/stromal cells for the production of new skin substitutes. *Stem Cells*. 2008;**26**(10):2713-2723
- [77] Chan RK et al. Development of a vascularized skin construct using adipose-derived stem cells from debrided burned skin. *Stem Cells International*. 2012;**2012**:1-11
- [78] Kellar BER, Diller RB, Machula H, Muller J. Biomimetic skin substitutes created from tropoelastin help to promote wound healing. *Frontiers in Bioengineering and Biotechnology*. DOI: 10.3389/conf.FBIOE.2016.01.00174

